

MANGROVE FUNGI OF KARACHI, PAKISTAN

FATIMA S. MEHDI*

S. M. SAIFULLAH*

SUMMARY : Fourteen different taxa of fungi were isolated from different parts of black mangrove Avicennia marina growing in Korangi Creek and Clifton areas of Karachi. They were identified as Alternaria maritima, Aspergillus flavus, Aspergillus niger, A. sulfurus, Aureobasidium pullulans, Bispora sp., Botrytis sp., Cladosporium sp., Humicola sp., Mucor sp., Penicillium sp., Phoma sp., Pythium sp. and Rhizopus sp. The most diverse group of them all was Deuteromycotina and the most frequent and easily grown Zygomycotina.

Key words : Fungi, aspergillus niger, aspergillus flavus.

INTRODUCTION

Mangroves were once neglected plants, but now they are gaining great importance because of the recent realization of their economic impact on the fishery resources of the area. Infact, the entire shrimp fishery of Pakistan, with more than a billion rupees in hard currency, depend upon the food and refuge provided by these plants. The mangroves of Pakistan have long been over looked but now their studies have just started (15-17) and emphasis is being placed more on ecology. There is now a tendency to study mangroves as an ecosystem and as such all related living and non-living components are being considered. Fungi also make a very important part of the ecosystem along with other microbes in turnover of the biomass (6), but unfortunately, they have received very little attention. Recently Hyde and Jones (2, 3), Jones and Hyde (6), Jones and Kuthbutheen (7), Kohlmeyer (9) and Venkatesan and Natarajan (19, 20) described fungi growing on mangroves from different parts of the world but information of Manglicolous fungi of Pakistan remains so far non existent. The present study was therefore carried out to fill in this important lacuna existing in the mangroves ecosystem studies of the area.

MATERIAL AND METHODS

Water samples, fresh leave, bark and seeds of the mangrove Avicennia marina were collected from a number of sites in Korangi Creek and Clifton area of Karachi, Pakistan (Figure 1) during the period July to September 199. Water samples were collected in sterile 150 ml screw cap flasks in replicates and the mangrove parts were placed in sterile polythene bags and either plated out immediately upon return to the laboratory or stored overnight in a cool incubation chamber at 4°C and the remainder at 18°C.

Isolation of black mangrove mycoflora

Bark and Seeds: Mangrove's bark and seeds were washed thoroughly in sterile sea water and cut into small pieces (2-3 mm²). These were then washed in sterile sea water containing penicillin and streptomycin at the rate of 2000 units to 1000 ml to suppress bacterial growth and placed on sea water agar, or Vitamin B₁ agar to which 1.5 g/l penicillin G and 1.5 g/l streptomycin sulphate had already been added. The surface of the plates were flooded with sterile sea water and the plates were incubated at 20°C. After 24 hours the plates were examined microscopically and any developing colonies were aseptically removed and inoculated on to fresh agar plates. After several successive plating pure cultures were obtained.

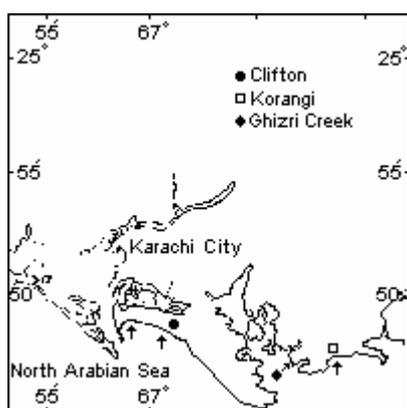
Phylloplane mycoflora of leaf

Plating of Leaf Discs: The initial leaf fungal population was determined by punching out five discs of leaf tissue from the

* From Department of Botany, University of Karachi, Karachi-75270, Pakistan.

leaves. The discs were cut out with a sterilized cork borer of 5 mm diameter, washed in five changes of 15 ml sterile sea water and then placed aseptically on the surface on WA plates at the rate of one disc per dish. The total number of disc taken was 15. The cultures were incubated at 20°C in the dark and examined microscopically at two to three days intervals, over a period of four to five weeks. Any fungi appearing were sub-cultured on to plates of PDA, CDA and CMA to encourage growth and sporulation and thus allowing identification to be made.

Figure 1: Map of the area of study.



RESULTS AND DISCUSSION

In all seventeen different species of micro-fungi were isolated from the water and mangroves of the two seaside localities (Figure 1, Tables 1 and 2). They were all secondary marine fungi (10), that is basically terrestrial forms. The species composition was similar to that found in other estuarine waters (1,13). Like the Indian Manglicolous fungi, Deuteromycotina was also the most diverse group in the area (19, 20). However, it was a different case in Hawaii where Kohlmeyer and Kohlmeyer (11) found Ascomycotina the most dominant group. Zygomycotina was also the most easily grown group in culture. In general the species composition of the area was limited in number as compared to other areas (3, 7,11), and this discrepancy may be due to the limited range of methodology used and the sampling in the present area.

Some differences in terms of species composition were noted between the two localities studies. Thus, all the 14 species mentioned in Table 2 were present in Clifton area whereas only eight in Korangi area. The species that were exclusive to the former area were *Alternaria marina*, *Aureobasidium pullulans*, *Cladosporium sp.*, *Humicola sp.*, *Phoma sp.* and *Pythium sp.* Difference in species composition was also noted in

different parts of mangroves. Thus maximum number of species were recorded on leaves whereas almost equal proportions from bark and seeds during the entire period of study in both places (Figures 2, 3, and 4). The difference parts with reference to locality. Thus, the leaves in Clifton areas allowed almost twice as many species to grow on them as in Korangi Creek area. Richness in species diversity in Clifton area may be due to the fact it is situated at the point of discharge of city's sewage and therefore is rich in organic matter.

Table 1: Micro-fungi isolated from water samples of Korangi Creek and Clifton (+ Indicates that a species was isolated at least once during the sampling period).

Fungal Species	Korangi Creek	Clifton
<i>Alternaria maritima</i> (Suth)	-	+
<i>Aspergillus flavus</i> Link	+	+
<i>A. niger</i> Vantieghem	+	-
<i>A. sulphurus</i> (Press.)		
Thom and Church	+	-
<i>Aureobasidium pullulans</i>	-	+
(Debary) Arnaud		
<i>Bispora</i> sp.	+	-
<i>Botrytis</i> sp.	+	+
<i>Cladosporium</i> sp.	-	+
<i>Fusarium solani</i> (Martius)	-	+
Appel and Wollen Weber		
<i>Humicola</i> sp.	-	+
<i>Mucor</i> sp.	+	-
<i>Penicillium</i> sp.	+	-
<i>P. expansum</i> Link Thom.	+	+
<i>P. brefeldianum</i> Dodge	+	+
<i>Phoma</i> sp.	-	+
<i>Pythium</i> sp.	-	+
<i>Rhizopus</i> sp.	+	+
Total number of species isolated	10	12

Table 2: Percentage frequency of fungal species isolated from leaf, bark and seed of black mangrove, collected from Korangi Creek and Clifton.

Fungal Species	JULY						AUGUST						SEPTEMBER						
	K			C			K			C			K			C			
	L	B	S	L	B	S	L	B	S	L	B	S	L	B	S	L	B	S	
<i>Alternaria maritima</i> (Suth)	-	-	-	30	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Aspergillus flavus</i> Link	-	20	25	10	40	-	10	-	40	25	-	-	-	20	-	20	-	10	
<i>A. niger</i> Vantieghem	20	-	-	-	-	-	25	10	20	20	20	-	30	20	10	10	-	-	
<i>A. sulphurus</i> (Press.)	40	-	-	-	-	-	40	10	10	10	-	-	10	15	-	40	40	-	
Thom and Church	-																		
<i>Aureobasidium pullulans</i>	-	-	-	40	-	-	-	-	-	50	-	-	-	-	-	60	30	-	
(Debary) Arnaud	-																		
<i>Bispora</i> sp.	-	20	-	40	-	-	10	8	-	25	20	-	30	20	20	40	-	-	
<i>Botrytis</i> sp.	50	40	-	50	-	-	10	15	-	-	-	10	10	-	40	35	-	-	
<i>Cladosporium</i> sp.	-	-	-	35	-	-	-	-	-	10	10	-	-	-	-	60	-	-	
<i>Humicola</i> sp.	-	-	-	20	-	-	-	-	-	-	-	20	-	-	-	-	-	20	
<i>Mucor</i> sp.	20	-	-	30	-	20	40	-	30	22	-	-	10	-	-	20	10	15	
<i>Penicillium</i> sp.	25	30	30	-	60	-	45	25	-	20	-	20	20	20	10	10	-	15	
<i>Phoma</i> sp.	-	-	-	15	40	10	-	-	-	10	15	-	-	-	-	-	10	-	
<i>Pythium</i> sp.	-	-	-	10	-	-	-	-	-	10	-	30	-	-	-	-	-	-	
<i>Rhizopus</i> sp.	50	40	-	35	-	10	10	10	-	-	-	-	-	10	10	-	-	-	
Total number of species isolated	6	5	2	11	3	3	8	6	4	10	4	4	6	6	5	9	4	4	

K:Korangi Creek; C:Clifton; L:Leaf; B:Bark; S:Seed

Figure 2

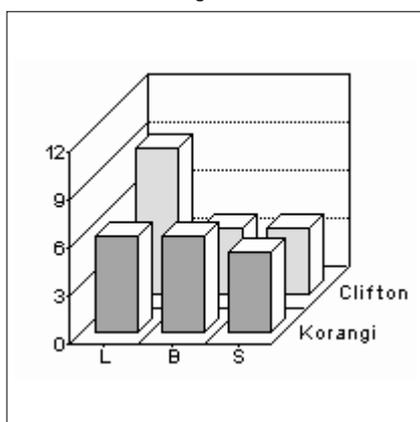


Figure 3

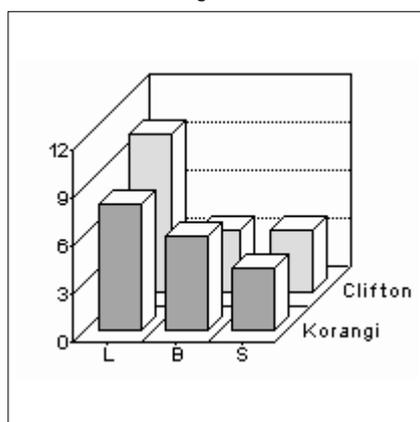
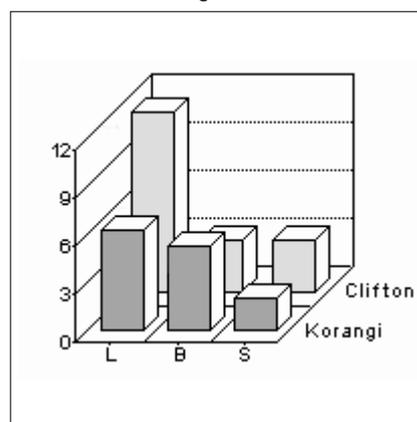


Figure 4



Figures 2, 3, 4: Relative composition of different species of mangrove fungi from Karachi during July (Figure 2), August (Figure 3) and September (Figure 4).

REFERENCES

1. Haythorn JM, Gareth EB, Harrison JL : Observations on marine algicolous fungi, including the Hyphomycete *Sigmoidea marina* sp nov. *Trans Br Mycol Soc*, 74:615-623, 1980.
2. Hyde KD, Jones EBG: Marine fungi from the Sychelles. VII. *Bathyascus grandispore* sp nov from Mangrove Wood. *Botanica Marina*, 30:413-416, 1987.
3. Hyde KD, Jones EBG : Marine Mangrove Fungi. *PSZNL. Marine Biology*, 9:15-33, 1988.
4. Hyde KD, Jones EBG : Intertidal Mangrove fungi from Brunei: *Lauto sporagigantea* gen et sp nov. *Botanica Marina*, 32:479-482, 1989.
5. Jones EBG, et al : Fungi on driftwood collected in the intertidal zone from the Phillippines. *Asian Marine Biology*, 5:103-106, 1988.
6. Jones EBG, Hyde KD : Methods for the study of Mangrove Marine Fungi. In: *Mangrove Microbiology; Role of Microorganisms in Nutrient Cycling of Mangrove Soils and Waters*, Ed by AD Agate, CV Subramanian, H Vannucci, pp 9-27, UUNDP, 1988.
7. Jones EBG, Kuthbutheen AJ : Malaysian Mangrove Fungi. *Sydowia*, 41:160-169, 1989.
8. Jones EBG, Hyde KD : Observations on poorly known Mangrove Fungi and a nomenclatural correction. *Mycotaxon*, 37:197-201, 1990.
9. Kohlmeyer J, Schatz S : *Aigialus* Gen Nov (Ascomycetes) with two new marine species from mangroves. *Trans Brit Mycol Soc*, 85:699-707, 1985.
10. Kohlmeyer J : Taxonomic studies of the Marine Ascomycotina in the Biology of Marine Fungi, Ed by ST Moss, Cambridge University Press, Cambridge.
11. Kohlmeyer J, Volkmann-Kohlmeyer B : Hawaiian marine fungi, including two new genera of Ascomycotina. *Mycol Res*, 92:410-421, 1989.
12. Raghu-Kumar S : *Thranstochytrium benthicola* sp nov. A new marine fungus from the north sea. *Trans Brit Mycol Soc*, 74:607-614, 1980.
13. Raghu-Kumar S : *Schizochytrium octosporium* sp nov and other thranstochytrids from the north sea (Roford Norway). *Trans Brit Mycol Soc*, 90:273-278, 1988.
14. Kagh-Kumar S : *Schizochytrium mangrovei* sp nov. A thranstochytrid from mangroves in India. *Trans Brit Mycol Soc*, 90:627-631, 1988.
15. Saifullah SM : Mangrove ecosystem of Pakistan, pp 69-80. In: *The Third Research on Mangroves in Middle East. Japan Cooperation Center for the Middle East. Publication No 137, Tokyo, 1982.*
16. Saifullah SM : Management of Mangroves in Pakistan, pp 7-9. In *Souvenir, National Seminar on "Study and Management in Coastal Zones in Pakistan". 23-26 September, 1991. Karachi, MRCC. Karachi University and UNESCO.*
17. Saifullah SM : Future prospects of Mangroves in development of Makran coast. In: *Proceedings of National Conference on Problems and Resources of Makran coast and plan of action for its Development. 28-30 September, 1991, Quetta (In press).*
18. Tan TK, Leong WF, Jones EBG : Succession of fungi on wood of *Avicennia alba* and *A. lanata* in Singapore. *Can J Bot*, 67:2687-2691, 1988.
19. Venkatesan T, Natarajan R : Intertidal mycoflora of Pichavaran mangroves near Porto Novo. *The Mangroves. Proc Nat Symp Biol Util Cons Mangroves*, Nov, pp 163-169, 1985.
20. Vankatesan T, Natarajan R : Rhizosphere mycoflora of Pichavaran mangroves near Porto Novo. *Proc All India Symp Man Plants Dona Paula GOA India*, pp 216-223, 1985.

Correspondence:
 Fatima S. Mehdi
 Department of Botany,
 University of Karachi,
 Karachi-75270,
 PAKISTAN.