



# Therapeutic Effects of Vitamin D<sub>3</sub> on Motor Functions Following Experimental Spinal Cord Injury

## *Deneysel Spinal Kord Yaralanmasında Vitamin D<sub>3</sub>'ün Motor Fonksiyonlar Üzerine Olan İyileştirici Etkisi*

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### Summary

**Objective:** Spinal cord injuries are frequently encountered and is a major health problem due to the severe disability they cause. The neurological damage of acute spinal cord trauma triggers primary mechanisms of injury and cell death activation cascades leading to tissue damage. The aim of this study was to investigate the therapeutic effects of vitamin D<sub>3</sub> in a rat model of spinal cord injury.

**Materials and Methods:** The study was performed at Haydarpaşa Numune Training Hospital, İstanbul, Turkey. Twenty-one Sprague-Dawley rats were randomly assigned to the following study groups: the control group, undergoing laminectomy procedure; trauma group, undergoing spinal cord injury after laminectomy; and trauma plus vitamin D<sub>3</sub> group, undergoing spinal cord injury after laminectomy and subsequent administration of vitamin D<sub>3</sub> on days 1, 3, 5, 7 intraperitoneally with a dosage of 1 mcg/kg/day. The functional outcome was evaluated by using inclined plane test and Drummond and Moore motor function score on days 1, 7, 14, 21. Spinal cord samples of the control, trauma and trauma plus vitamin D<sub>3</sub> groups were obtained for histopathologic evaluations after clinical examinations and were examined under light microscope.

**Results:** Inclined plane test scores and motor function scores of vitamin D<sub>3</sub> group were significantly higher than the trauma group at 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> weeks.

**Conclusion:** In our study we have investigated the therapeutic effects of vitamin D<sub>3</sub> and found out that it has a positive impact to the recovery process of the spinal cord injury. (Turkish Journal of Neurology 2015; 21:55-61)

**Key Words:** Experimental spinal cord injury, antioxidant treatment, vitamin D<sub>3</sub>

### Özet

**Amaç:** Akut spinal kord yaralanması (SKY) sonrası nörolojik hasar, primer mekanik yaralanma ile olduğu kadar sekonder olarak doku hasarına yol açan hücre ölümü aktivasyon kaskatlarından da kaynaklanmaktadır. Bu çalışmada vitamin D<sub>3</sub>'ün sıçanlarda oluşturulan deneysel SKY sonrasında motor fonksiyonlar üzerindeki iyileştirici etkinliği araştırıldı.

**Gereç ve Yöntem:** Bu çalışma Sağlık Bakanlığı Haydarpaşa Numune Eğitim ve Araştırma Hastanesi'nde yapıldı. Çalışmada 3 grupta, 7'şer adet olmak üzere toplam 21 adet Sprague-Dawley cinsi sıçan kullanıldı. İlk gruba sadece laminektomi yapıldı (kontrol grubu). İkinci gruba laminektomi yapıp omurilik yaralanması oluşturuldu (travma grubu). Üçüncü gruba laminektomi yapıp omurilik yaralanması oluşturuldu ve 1., 3., 5., 7. günlerde intraperitoneal olarak 1 mcg/kg/gün dozunda vitamin D<sub>3</sub> enjeksiyonu yapıldı (vitamin D<sub>3</sub> grup). Deneklerin fonksiyonel iyileşmeleri cerrahi işlem sonrası 1., 7., 14. ve 21. günlerde eğik düzlem testi, Drummond ve Moore motor fonksiyon skoru ile değerlendirildi. Kontrol, travma ve vitamin D<sub>3</sub> uygulanan gruplardaki doku parçaları hematoksil-eozin (HE) boyası ile boyanıp ışık mikroskopunda incelendi.

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**Bulgular:** Vitamin D<sub>3</sub> grubunda eğik düzlem testi sonuçları ve motor fonksiyon skorları travma grubu ile karşılaştırıldığında 1., 2. ve 3. haftalarda anlamlı olarak daha yüksek bulundu.

**Sonuç:** Bu çalışmamızda, sıçanlarda oluşturulan deneysel SKY'de vitamin D<sub>3</sub>'ün tedavi edici etkisi araştırılmış olup klinik olarak iyileşme sürecine anlamlı etkinliği olduğu saptanmıştır. (Türk Nöroloji Dergisi 2015; 21:55-61)

**Anahtar Kelimeler:** Deneysel spinal kord yaralanması, antioksidan tedavi, vitamin D<sub>3</sub>

## Introduction

Today, spinal cord injury (SPI) remains as an important topic due to its incidence rate, the magnitude of physical, psychosocial and economic damages it incurs, and the lack of universally accepted treatment protocols. Recent attempts at developing pharmacological treatment methods highlighted the importance of understanding the pathophysiology of the processes following the trauma (1). Even though there is complete loss of function following primary or mechanical trauma in spinal cord injury cases, it is rarely a reason for total transection. It is also known that the biochemical and pathological changes in the cord can worsen the damage (2). Hypoxia on the cord can be seen in the early stage of acute primary injury. Following the primary injury of the tissue, a secondary damage stage involving hypoxia-induced electrolyte imbalance, neural excitation, glutamate release, formation of free radicals and inflammation progresses (3,4,5). In the secondary injury, the cascade of events that take place is a product of the activation of endogenous cell-death pathways. Despite the promising therapeutic effects of many agents, only methylprednisolone was shown to be effective in treatment in large-scale clinical studies (6).

Vitamin D is not really a vitamin but a secosteroid hormone and it has two types that come from different sources but have similar structure and composition. Ergocalciferol (vitamin D<sub>2</sub>) and cholecalciferol (vitamin D<sub>3</sub>) can be absorbed from nutrients but the main source of vitamin D is first synthesized on the skin with UVB rays and then reaches its active form after being hydroxylated in the liver and kidneys. In liver, vitamin D<sub>3</sub> turns into its active form calcidiol and vitamin D<sub>2</sub> turns into its active form 25(OH)D<sub>2</sub> (7,8,9). The biological functions of Vitamin D<sub>3</sub> include the regulation of calcium balance, cell differentiation in the nervous system, release of neutrophils and the activation of the enzymes and genes that play key roles in neurotransmitter synthesis (10). Brain is a target tissue for the effects of vitamin D<sub>3</sub>. It was shown that vitamin D receptor (VDR) which is a member of the nuclear hormone receptor superfamily is localized in brain, spinal cord and glial cells (11,12). In addition, the genes coding the enzymes associated with vitamin D<sub>3</sub> metabolism are expressed by the central nervous system (CNS) cells. The widespread distribution of VDR in different regions of the sensory, motor and limbic systems suggests that vitamin D<sub>3</sub> may possess many additional functional properties (13).

Experimental neurodegeneration models suggested vitamin D<sub>3</sub> to be neuroprotective (14,15,16,17). There is scientific evidence suggesting that vitamin D<sub>3</sub> also regulates immune system and brain functions (18). Studies showed that through VDR mediation vitamin D<sub>3</sub> prevents cytotoxicity in cortical

neurons by regulating inducible nitric oxide synthesis (iNOS) and it inhibits the iNOS expression in macrophage, active microglia and astrocytes (19,20). Another study showed that the blocking of iNOS through selective inhibitors decreases blood-spinal cord barrier damage induced by spinal injury, edema formation and cell reactions, therefore imposing a restorative influence over motor functions (21,22). Based on the neuroprotective efficacy of the pharmacological analogues of vitamin D<sub>3</sub> in neurodegenerative and neuroimmune diseases, this study looked at the restorative effects of vitamin D<sub>3</sub> on neurological functions in experimentally induced spinal cord injuries.

## Materials and Methods

This study was conducted in Ministry of Health Haydarpaşa Numune Training and Research Hospital under the approval of animal rights ethical board. Three groups of 7 Sprague-Dawley rats (21 in total) were used in the study. The weights of the rats in each group were between 180-220. Group 1 (n=7 rats): Laminectomy-only (control) group, group 2 (n=7 rats): Laminectomized and spinal cord trauma-induced group (traumatized group), group 3 (n=7 rats): Laminectomized, spinal cord traumatized and vitamin D<sub>3</sub>-administered group (on the 1<sup>st</sup>, 3<sup>rd</sup>, 5<sup>th</sup>, and 7<sup>th</sup> days, intraperitoneally, once a day 1 mcg/kg/day of Calcijex amp. 1 mcg/1 mL, Abbott) (vitamin D<sub>3</sub> group) (23).

For general anesthesia, 2 mg/kg ketamine HCl (Ketalar, Parke-Davis Eczacıbaşı-İstanbul) was given intraperitoneally before the surgical operation. Rats were positioned on special boards in prone position to enable comfortable access the during surgical operation. Thoracic regions were first sterilized with PVD iodine and then shaved. The region was sterilized once more with PVD iodine after shaving. Body temperature was fixed at 37 °C during the operation until the anesthetic effect wore off. Taking the interscapular distance as a reference, a 2 cm incision at the level of T5-T12 was made which penetrated dermis, epidermis and cleared away paravertebral muscles, eventually revealing the laminae. After the T7-8-9 laminectomy, standard spinal cord injury was induced by using modified Allen weight-drop model (24). Following hemostasis, primary suturing was applied on paravertebral muscles and skin using 3/0 vicryl in a way that preserved the anatomical layering. The rats were woken normally in room temperature. In the study, the animals that received T 7-8-9 total laminectomy all had intact dura matter. Using the modified Allen weight-drop model, SCI was induced by dropping a 10 g weight through a 10 cm long glass tube with 0.5 diameter. The rats that were given SCI were woken up in room temperature and their motor functions were examined. Fourteen rats on which trauma was induced (the trauma and

vitamin D<sub>3</sub> groups) were seen to be paraplegic. Seven rats that received laminectomy only (control group) all retained their full motor strength. Rats were placed in individual cages that were kept in room temperature and were given standard rat food. Paraplegic 14 rats were fed using orogastric tube during the early post-op stage. Bladder functions were monitored using the urinary catheters. Vitamin D<sub>3</sub> (Calcijex 1 mcg/1 mL) was acquired from Abbott Pharmaceuticals (İstanbul). Seven rats in the third group that had laminectomy and spinal injury using the weight-drop model received 1 mcg/kg/day vitamin D<sub>3</sub> intraperitoneally once a day on the 1<sup>st</sup>, 3<sup>rd</sup>, 5<sup>th</sup> and 7<sup>th</sup> days.

#### Evaluation of Functional Recovery

a) **Inclined plane test (25):** Functional recovery of the rats were evaluated by the inclined plane test developed by Rivliv and Tator which is commonly used in experimental acute spinal cord injury. Rats in all three groups went under inclined plane test on the 1<sup>st</sup>, 7<sup>th</sup>, 14<sup>th</sup> and 21<sup>st</sup> days following the surgery.

b) **Clinical motor exam:** The rats in the study went under motor examination in order to assess the functional recovery. The motor functions of the rats were evaluated on the 1<sup>st</sup>, 7<sup>th</sup>, 14<sup>th</sup> and 21<sup>st</sup> days following the surgery with the inclined plane test using Drummond and Moore criteria (26). Inclined plane test and clinical motor exam results were compared between groups, and within groups on the 1<sup>st</sup>, 7<sup>th</sup>, 14<sup>th</sup> and 21<sup>st</sup> days.

#### Histopathological Examination

Light microscopy examinations in our study were conducted in Ministry of Health Haydarpaşa Numune Training and Research Hospital Pathology Laboratory. At the end of 3 weeks, rats were anesthetized using 2 mg/kg intraperitoneal ketamine HCl and put in a supine position. After the anesthesia, the old incisions were reopened and the 30 mm spinal cord segments at the level of T 7-8-9 were examined under light microscopy. All of the tissue segments of control, trauma and vitamin D<sub>3</sub> groups were

examined. Following the routine tissue examination, paraffin blocks were cut in 5-micron thickness and dyed with hematoxylin-eosin (HE) before being examined under the light microscope.

#### Statistical Analysis

The data was analyzed using SPSS (Statistical Package for Social Sciences) for Windows 10.0. The quantitative data in the study did not follow a normal distribution. Therefore we used Kruskal Wallis test to compare the groups in terms of dependent variables, and Mann Whitney U test to identify the different group. Within-group tests were conducted using Wilcoxon signed test. The results were evaluated within 95% confidence interval and  $p < 0.05$  significance level.

## Results

#### Evaluation of the Inclined Plane Test Results

The levels of inclined plane test for each group on the 1<sup>st</sup> day, 1<sup>st</sup> week, 2<sup>nd</sup> week and 3<sup>rd</sup> week are shown in Table 1.

#### Rating of the Motor Functions (according to Drummond and Moore criteria)

The motor function scores for each group on the 1<sup>st</sup> day, 1<sup>st</sup> week, 2<sup>nd</sup> week and 3<sup>rd</sup> week are shown in Table 1.

#### Light Microscopy

The spinal cord cross-section of the control group, which did not receive trauma following laminectomy, is shown in Figures 1 and 2. In the trauma group cross-sections, there were liquefactive necrosis causing scattered cavities, dense histiocytes, and polymorphic leukocyte infiltration. Additionally, the surrounding tissue was infarction-like with axonal swelling and sparse axonal spheroids (Figures 3 and 4). The extent of the damage in vitamin D<sub>3</sub> group cross-sections covered a larger area than the trauma group, which did not receive the medication, and the morphological changes in the damaged region were at least as severe as the trauma group (Figures 5 and 6).

Table 1. Evaluation of the inclined plane degrees of the groups on the 1<sup>st</sup> day, 1<sup>st</sup>, 2<sup>nd</sup>, and 3<sup>rd</sup> weeks

Inclined Plane	Group 1 (Control) Mean ± SD (Median)	Group 2 (Trauma) Mean ± SD (Median)	Group 3 (Vitamin D <sub>3</sub> ) Mean ± SD (Median)	P
1 <sup>st</sup> day	60.00±0.00 (60)	10.00±0.00 (10)	10.00±0.00 (10)	0.001**
1 <sup>st</sup> week	60.00±0.00 (60)	11.43±2.44 (10)	13.57±2.44 (15)	0.001**
2 <sup>nd</sup> week	60.00±0.00 (60)	14.28±3.45 (15)	22.86±2.67 (25)	0.001**
3 <sup>rd</sup> week	60.00±0.00 (60)	25.71±3.45 (25)	42.86±14.54 (45)	0.001**

Kruskal Wallis Test, \*\*p<0.01

Table 2. Evaluation of the motor function of the groups on the 1<sup>st</sup> day, 1<sup>st</sup>, 2<sup>nd</sup>, and 3<sup>rd</sup> weeks

Motor Fonksiyon Skorları	Group 1 (Control) Mean ± SD (Median)	Group 2 (Trauma) Mean ± SD (Median)	Group 3 (Vitamin D <sub>3</sub> ) Mean ± SD (Median)	P
1 <sup>st</sup> day	4.00±0.00 (4)	0.00±0.00 (0)	0.00±0.00 (0)	0.001**
1 <sup>st</sup> week	4.00±0.00 (4)	0.28±0.49 (0)	0.57±0.53 (1)	0.001**
2 <sup>nd</sup> week	4.00±0.00 (4)	1.00±0.76 (1)	1.50±0.41 (1.5)	0.001**
3 <sup>rd</sup> week	4.00±0.00 (4)	1.50±0.50 (1.5)	2.43±0.45 (2.5)	0.001**

Kruskal Wallis Test, \*\*p<0.01

## Discussion

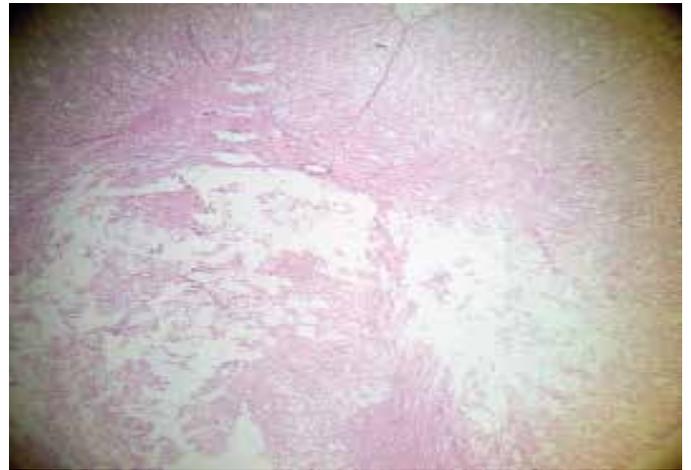
In the past 20 years, researchers have been focusing on the pathophysiological mechanisms of acute spinal cord injury in order to fix the neurological functions (27,28,29,30,31). Neurological damage following SCI is caused by the cell death activation cascades (secondary injury) as much as the 'primary mechanical injury' (32). Secondary spinal cord injury is the damage resulting from a series of pathophysiological processes triggered by the primary injury in the following hours or days (33,34,35). Numerous experimental studies showed a dose and severity dependent decrease in spinal cord blood-flow that gradually worsens following the injury (36). An interesting finding is that ischemia may be preventable if it is treated during the first few hours of the progressive decline in the post-traumatic ischemia (37,38). An important mechanism responsible for the advancement of the secondary damage is the increase in the NO (nitric oxide) synthesis (39,40). With the

increased concentration, NO becomes neurotoxic and facilitates the secondary damage as a free radical. Being one of the three enzymes playing a role in the nitric oxide synthesis, iNOS does not exist in healthy tissues but causes NO synthesis after being stimulated by the inflammatory mediators and cytokines during the pathological processes (41,42,43). NOS inhibition in the early stages of SCI provides the benefit of improving neurological functions and histopathological changes (44).

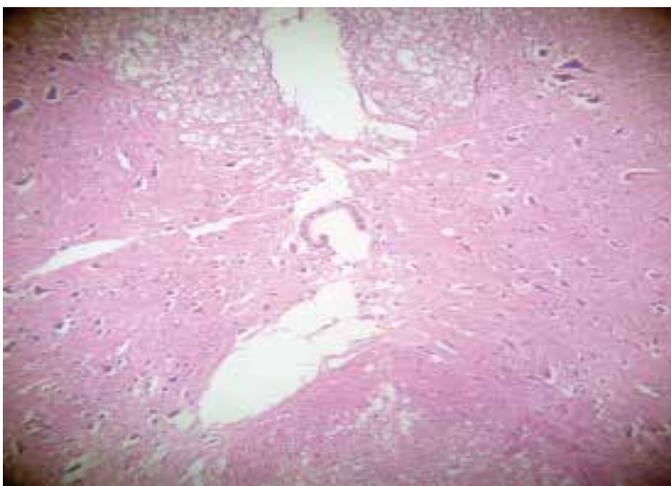
Even though it was initially assumed that calcitriol, the active form of vitamin D, was produced by the liver and kidneys, it was later discovered that many other organs including the brain also express vitamin D1 hydroxylase (45). Additionally, VDRs propagate over large areas of both human and rat brains (46,47). Studies showed that through VDR, vitamin D regulates brain-based neurotropic factor, nerve growth factor and the gene expression coding neurotrophin 3 (48). It was shown that addition of vitamin D<sub>3</sub> in cell cultures increases neurotrophin expression



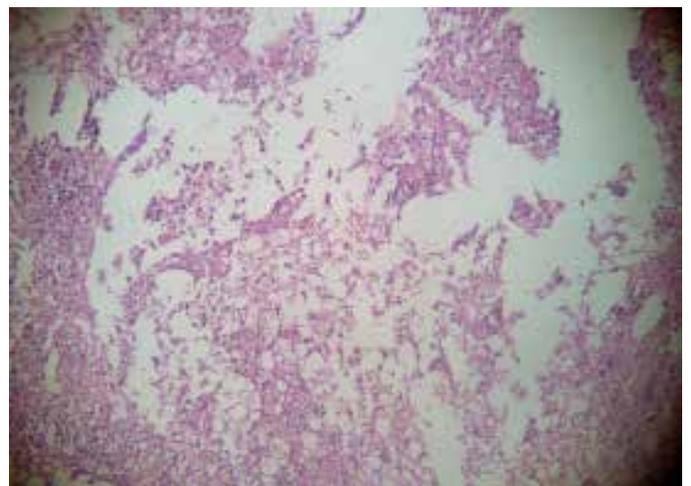
**Figure 1.** Spinal cord in the control group following laminectomy, without trauma (HE, x40 magnified)



**Figure 3.** Trauma group. Spinal cord following laminectomy and trauma (HE, x40)

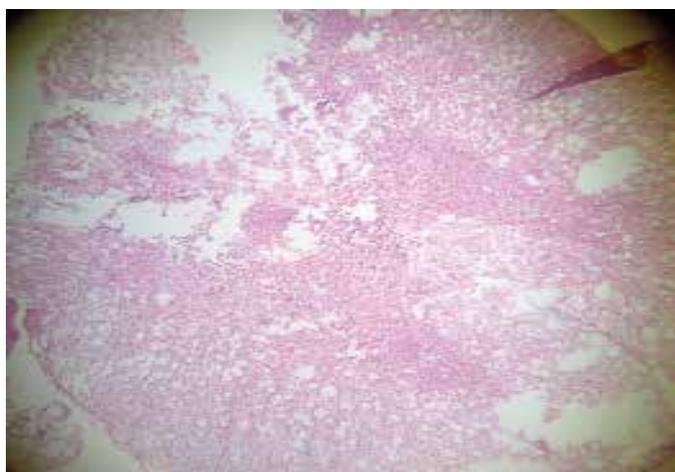


**Figure 2.** Control group (HE, x100)

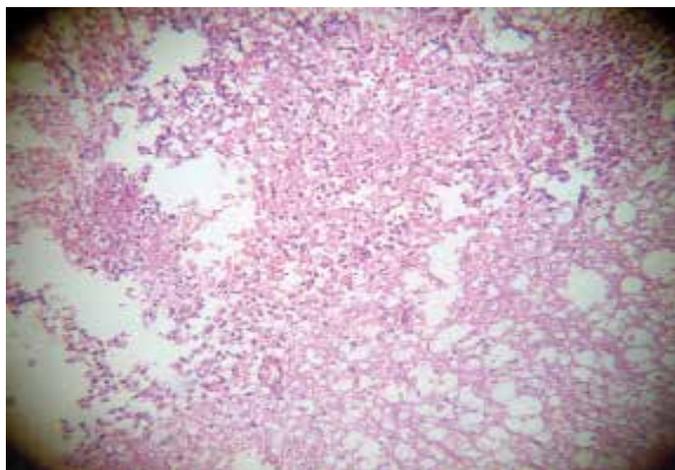


**Figure 4.** Trauma group (HE, x100)

and nerve growth (49). Another study found that neuronal growth factor expression decreases in the rat brain when vitamin D<sub>3</sub> concentration in the environment is decreased (50). Chabas et al. found that vitamin D (especially vitamin D<sub>3</sub>) plays an important role in axogenesis and myelination through the genes associated with calcitriol (51,52). In many experimental neurodegeneration models, vitamin D<sub>3</sub> was also shown to be neuroprotective (53,54,55). Studies have shown that vitamin D<sub>3</sub> decreases the formation of nitrite, reduces oxidative stress by increasing gamma glutamyl transpeptidase (56). In addition, in vivo studies showed that it decreases cortical infarcts after middle cerebral artery clogging (57). In the present study, which was motivated by these findings, our functional recovery measurements using inclined plane test showed that the drug group which received vitamin D<sub>3</sub> showed improvement following the first week. It was seen that the rats received vitamin D<sub>3</sub> and those that simply received the trauma had started to differ in statistically significant amounts after two



**Figure 5.** Traumatized, post-op on the 1<sup>st</sup>, 3<sup>rd</sup>, 5<sup>th</sup> and 7<sup>th</sup> days 1 mcg/ml dose vitamin D<sub>3</sub> group (HE, x40)



**Figure 6.** Traumatized and given 1 mcg/ml dose vitamin D<sub>3</sub>, post-op on the 1<sup>st</sup>, 3<sup>rd</sup>, 5<sup>th</sup> and 7<sup>th</sup> days (HE, x100)

weeks in terms of incline plane degrees. The analysis of motor function scores using Drummond and Moore criteria showed that the vitamin D<sub>3</sub> group showed improvement in motor functions after two weeks. A statistically significant difference in motor functions between the vitamin group and the trauma-only group developed on the third week. These findings were interpreted as vitamin D<sub>3</sub> inducing neuronal growth factors and contributing to axonogenesis and additionally facilitating the healing process through its antioxidant effect. Additionally another study showed an increase in the muscle mass and strength in hemiplegic patients following vitamin D treatment, which was interpreted as vitamin D treatment increasing physical performance. These results also explain the clinical improvement seen in our study (58,59).

Our light microscopy study showed regional cavities caused by liquefactive necrosis, dense histiocytes and polymorph leukocyte infiltration accompanied by infarction-like appearance in the surrounding tissues, axonal swelling and sporadic axonal steroids in the trauma group. Vitamin D<sub>3</sub> group's slices showed a similar appearance in the morphological damage area. The reason why the clinical improvement was not paralleled by the histology may be that the precise time frame for the destructive effects of the secondary injury is unknown. The effect of the damage can extend to as much as 3-6 weeks and it was assumed that these effects were still in progress during our study (60,61).

In this study, we investigated the neuroprotective effects of vitamin D<sub>3</sub> in SCI created by modified Allen weight-drop model. Even though there was clinical improvement in the neurological functions, microscopy studies did not show meaningful changes. Further studies are needed for the clinical use of vitamin D<sub>3</sub>.

**Ethical Approval:** The manuscript was derived from a thesis. Ethical approval for the study was given by the Ministry of Health Haydarpaşa Numune Training and Research Hospital Ethical Board

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## References

1. Sontag VKH. History of degenerative and traumatic diseases of the spine. In a history of neurosurgery. Greenblat SH. American Association of neurological Surgeons, Washington; 1997:355-357.
2. Faden AI, Simon RP. A potential role for excitotoxins in the pathophysiology of spinal cord injury. *Ann Neurol* 1988;23:623-626.
3. Croft TJ, Brodkey JS, Nulsen FE. Reversible spinal cord trauma: a model for electrical monitoring of spinal cord function. *J Neurosurg* 1972;36:402-406.

4. Delamarter RB, Sherman J, Carr JB. Pathophysiology of spinal cord injury. Recovery after immediate and delayed decompression. *J Bone Joint Surg Am* 1995;77:1042-1049.
5. Dumont AS, Dumont RJ, Oskouian RJ. Will improved understanding of the pathophysiological mechanisms involved in acute spinal cord injury improve the potential for therapeutic intervention? *Curr Opin Neurol* 2002;15:713-720.
6. Boyaci MG, Eser O, Kocogullari CU, Karavelioglu E, Tokyol C, Can Y. Neuroprotective effect of alpha-lipoic acid and methylprednisolone on the spinal cord ischemia/reperfusion injury in rabbits. *Br J Neurosurg* 2014;1-6.
7. Heaney RP, Weaver CM. Calcium and vitamin D. *Endocrinol Metab Clin North Am* 2003;32:181-194.
8. Brown AJ, Dusso A, Slatopolsky E. Vitamin D. *Am J Physiol* 1999;277:157-175.
9. Segaert S, Bouillon R. Vitamin D and regulation of gene expression. *Curr Opin Clin Nutr Metab Care* 1998;1:347-354.
10. Wang Y, Chiang YH, Su TP, Hayashi T, Morales M, Hoffer BJ, Lin SZ.: Vitamin D(3) attenuates cortical infarction induced by middle cerebral arterial ligation in rats. *Neuropharmacology* 2000;39:873-80.
11. Stumpf WE, Sar M, Clark SA, DeLuca HF. Brain target sites for 1,25 dihydroxyvitamin D3. *Science* 1982;215:1403-1405.
12. Stumpf WE, Clark SA, O'Brien LP, Reid FA. 1,25(OH)2 vit D3 sites of action in spinal cord and sensory ganglion. *Anat Embryol (Berl)* 1987;177:307-310.
13. Prüfer K, Veenstra TD, Jirikowski GF, Kumar R. Distribution of 1,25 dihydroxyvitamin D3 receptor immunoreactivity in the rat brain and spinal cord. *J Chem Neuroanat* 1999;16:135-145.
14. Orme RP, Bhargal MS, Fricker RA. Calcitriol imparts neuroprotection in vitro to midbrain dopaminergic neurons by upregulating GDNF expression. *PLoS One* 2013;8:62040.
15. Lin AM, Chen KB, Chao PL. Antioxydative effect of vitamin D3 on zinc-induced oxydative stres in CNS. *Ann N Y Acad Sci* 2005.;1053:319-329.
16. Jang W, Kim HJ, Li H, Jo KD, Lee MK, Song SH, Yang HO. 1,25-Dihydroxyvitamin D attenuates rotenone-induced neurotoxicity in SH-SY5Y cells through induction of autophagy. *Biochem Biophys Res Commun* 2014;451:142-147.
17. Wang JY, Wu JN, Cherg TL, Hoffer BJ, Chen HH, Borlongan CV, Wang Y. Vitamin D3 attenuates 6-hydroxydopamine-induced neurotoxicity in rats. *Brain Res* 2001;904:67-75.
18. Deluca HF, Cantorna MT. Vitamin D: its role and uses in immunology. *FASEB J* 2001;15:2579-2585.
19. Dursun E, Gezen-Ak D, Yilmazer S. A new mechanism for amyloid- induction of iNOS: vitamin D-VDR pathway disruption. *J Alzheimers Dis* 2013;36:459-474.
20. Tardivo V, Crobeddu E, Pilloni G, Fontanella M, Spena G, Panciani PP, Berjano P, Ajello M, Bozzaro M, Agnoletti A, Altieri R, Fiumefreddo A, Zenga F, Ducati A, Garbossa D. Say "no" to spinal cord injury: is nitric oxide an option for therapeutic strategies? *Int J Neurosci* 2015;125:81-90.
21. Genovese T, Mazzon E, Mariotto S, Menegazzi M, Cardali S, Conti A, Suzuki H, Bramanti P, Cuzzocrea S. Modulation of nitric oxide homeostasis in a Mouse model of spinal cord injury. *J Neurosurg Spine* 2006;4:145-153.
22. Garcion E, Nataf S, Berod A, Darcy F, Brachet P. 1,25-Dihydroxyvitamin D3 inhibits the expression of inducible nitric oxide synthase in rat central nervous system during experimental allergic encephalomyelitis. *Brain Res Mol Brain Res* 1997;45:255-267.
23. Fu J, Xue R, Gu J, Xiao Y, Zhong H, Pan X, Ran R. Neuroprotective effect of calcitriol on ischemic/reperfusion injury through the NR3A/CREB pathways in the rat hippocampus. *Mol Med Rep* 2013;8:1708-1714.
24. Allen AR. Surgery of experimental lesion of spinal cord equivalent to crush injury of fracture dislocation of spinal column. A preliminary report. *JAMA* 1911;57:878-880.
25. Rivlin AS, Tator CH. Objective clinical assessment of motor function after experimental spinal cord injury in the rat. *J Neurosurg* 1977;47:577-581.
26. Drummond JC, Moore SS. The influence of dextrose administration on neurologic outcome after temporary spinal cord ischemia in the rabbit. *Anesthesiology* 1989;70:64-67.
27. Zhou X, He X, Ren Y. Function of microglia and macrophages in secondary damage after spinal cord injury. *Neural Regen Res* 2014;9:1787-1795.
28. Ramer LM, Ramer MS, Bradbury EJ. Restoring function after spinal cord injury: towards clinical translation of experimental strategies. *Lancet Neurol* 2014;13:1241-1256.
29. Zhang J, Wei H, Lin M, Chen C, Wang C, Liu M. Curcumin protects against ischemic spinal cord injury: The pathway effect. *Neural Regen Res* 2013;8:3391-3400.
30. Naseem M, Parvez S. Role of melatonin in traumatic brain Injury and spinal cord injury. *Scientific World Journal* 2014;2014:586270.
31. Tator CH. Update on the pathophysiology and pathology of acute spinal cord injury. *Brain Pathol* 1995;5:407-413.
32. Emery E, Aldana P, Bunge MB, Puckert W, Srinivasan A, Keane RW, Bethea J, Levi AD. Apoptosis after traumatic human spinal cord injury. *J Neurosurg* 1998;89:911-920.
33. Tator CH, Koyanagi I. Vascular mechanisms in the pathophysiology of human spinal cord injury. *J Neurosurg* 1997;86:483-492.
34. Kasinathan N, Vanathi MB, Subrahmanyam VM, Rao JV. A Review on response of immune system in spinal cord injury and therapeutic agents useful in treatment. *Curr Pharm Biotechnol* 2015;16:26-34.
35. Hall ED, Wang JA, Bosken JM, Singh IN. Lipid peroxidation in brain or spinal cord mitochondria after injury. *J Bioenerg Biomembr* 2015.
36. Dohrman GJ, Wagner FC Jr, Bucy PC. The microvasculature in transitory traumatic paraplegia. An electron microscopic study in the monkey. *J Neurosurg* 1971;35:263-271.
37. Dumont RJ, Okonkwo DO, Verma S, Hurlbert RJ, Boulos PT, Ellegala DB, Dumont AS. Acute spinal cord injury, Part 1: pathophysiologic mechanisms. *Clin Neuropharmacol* 2001;24:254-264.
38. Amar PA, Levy ML. Pathogenesis and pharmacological strategies for mitigating secondary damage in acute spinal cord injury. *Neurosurgery* 1999;44:1027-1039.
39. Koozekanani SH, Vise WM, Hashemi RM, McGhee RB. Possible mechanisms for observed pathophysiological variability in experimental spinal cord injury by the method of Allen. *J Neurosurg* 1976;44:429-434.
40. Isaksson J, Faroog M, Olsson Y. Improved functional outcome after spinal cord injury in iNOS-deficient mice. *Spinal cord* 2005;43:167-170.
41. Díaz-Ruiz A, Ibarra A, Pérez-Severiano F, Guízar-Sahagún G, Grijalva I, Ríos C. Constitutive end inducible nitric oxide synthase activities after spinal cord contusion in rats. *Neurosci Lett* 2002;319:129-132.
42. Liu C, Jin A, Zhou C, Chen B. Nitric oxide synthase gene expression in injured spinal cord tissue. *Chin Med J* 2002;115:740-742.
43. Sharma HS, Badgaiyan RD, Alm P, Mohanty S, Wiklund L. Neuroprotective effects of nitric oxide synthase inhibitors in spinal cord injury-inducible pathophysiology and motor functions: an experimental study in the rat. *Ann N Y Acad Sci* 2005;1053:422-434.
44. Jiang Y, Gong FL, Zhao GB, Li J. Chrysin suppressed inflammatory responses and the inducible nitric oxide synthase pathway after spinal cord injury in rats. *Int J Mol Sci* 2014;15:12270-12279.
45. Eyles DW, Smith S, Kinobe R, Hewison M, McGrath JJ. Distribution of the vitamin D receptor and 1 alpha-hydroxylase in human brain. *J Chem Neuroanat* 2005;29:21-30.
46. Langub MC, Herman JP, Malluche HH, Koszewski NJ. Evidence of functional vitamin D receptors in rat hippocampus. *Neuroscience* 2001;104:49-56.
47. Veenstra TD, Prüfer K, Koenigsberger C, Brimijoin SW, Grande JP, Kumar R. 1,25-Dihydroxyvitamin D3 receptors in the central nervous system of the rat embryo. *Brain Res* 1998;804:193-205.
48. Wang TT, Tavera-Mendoza LE, Laperriere D, Libby E, MacLeod NB, Nagai Y, Bourdeau V, Konstorum A, Lallemand B, Zhang R, Mader S, White JH. Large-scale in silico and microarray-based identification of direct 1,25-dihydroxyvitamin D3 target genes. *Mol Endocrinol* 2005;19:2685-2695.
49. Marini F, Bartoccini E, Cascianelli G, Voccoli V, Baviglia MG, Magni MV, Garcia-Gil M, Albi E. Effect of 1alpha, 25-dihydroxyvitamin D3 in embryonic hippocampal cells. *Hippocampus* 2010;20:696-705.
50. Féron F, Burne TH, Brown J, Smith E, McGrath JJ, Mackay-Sim A, Eyles DW. Developmental Vitamin D3 deficiency alters the adult rat brain. *Brain Res Bull* 2005;65:141-148.
51. Chabas JF, Stephan D, Marqueste T, Garcia S, Lavaut MN, Nguyen C, Legre R, Khrestchatsky M, Decherchi P, Féron F. Cholecalciferol (vitamin D) improves myelination and recovery after nerve injury. *PLoS One* 2013;8:65034.

52. Chabas JF, Alluin O, Rao G, Garcia S, Lavaut MN, Risso JJ, Legre R, Magalon G, Khrestchatsky M, Marqueste T, Decherchi P, Feron F. Vitamin D2 potentiates axon regeneration. *J Neurotrauma* 2008;25:1247-1256.
53. Malcok UA, Sengul G, Kadioglu HH, Aydin IH. Therapeutic effects of vitamin D3 in a rat diffuse axonal injury model. *J Int Med Res* 2005;33:90-95.
54. Javidan AN, Sabour H, Latifi S, Vafa M, Shidfar F, Khazaiepour Z, Shahbazi E, Rahimi A, Razavi SH. Calcium and vitamin D plasma concentration and nutritional intake status in patients with chronic spinal cord injury: A referral center report. *J Res Med Sci* 2014;19:881-884.
55. Féron F, Marqueste T, Bianco J, Gueye Y, Chabas JF, Decherchi P. Repairing the spinal cord with vitamin D: a promising strategy. *Biol Aujourd'hui* 2014;208:69-75.
56. Ibi M, Sawada H, Nakanishi M, Kume T, Katsuki H, Kaneko S, Shimohama S, Akaike A. Protective effects of 1 alpha,25-(OH)(2)D(3) against the neurotoxicity of glutamate and reactive oxygen species in mesencephalic culture. *Neuropharmacology* 2001;40:761-771.
57. Wang Y, Chiang YH, Su TP, Hayashi T, Morales M, Hoffer BJ, Lin SZ. Vitamin D(3) attenuates cortical infarction induced by middle cerebral arterial ligation in rats. *Neuropharmacology* 2000;39:873-880.
58. Sato Y, Iwamoto J, Kanoko T, Satoh K. Low-dose vitamin D prevents muscular atrophy and reduces falls and hip fractures in women after stroke: a randomized controlled trial. *Cerebrovasc Dis* 2005;20:187-192.
59. Cannell JJ, Hollis BW, Sorenson MB, Taft TN, Anderson JJ. Athletic performance and vitamin D. *Med Sci Sports Exerc* 2009;41:1102-1110.
60. Christie SD, Comeau B, Myers T, Sadi D, Purdy M, Mendez I. Duration of lipid peroxidation after acute spinal cord injury in rats and the effect of methylprednisolone. *Neurosurg Focus* 2008;25:5.
61. Kamencic H, Griebel RW, Lyon AW, Paterson PG, Juurlink BH. Promoting glutathione synthesis after spinal cord trauma decreases secondary damage and promotes retention of function. *FASEB J* 2001;15:243-250.