Analysis of late-onset neonatal sepsis cases in a level three neonatal intensive care unit

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ABSTRACT

OBJECTIVE: Newborns in neonatal intensive care units (NICUs) are at high risk for developing nosocomial infections (NIs), which may result in morbidity and mortality. In this study, we aimed to ascertain the bacteriological profiles and their antimicrobial susceptibility patterns in NIs.

METHODS: We conducted a cross-sectional study in the NICU. Late-onset sepsis (LOS) cases confirmed with blood cultures were evaluated retrospectively. Laboratory parameters, demographics, and clinical data were collected and analyzed from hospital records retrospectively.

RESULTS: Of the 1210 infants in the NICU, 76 (6.3%) had LOS. A total of 86 episodes of LOS were documented; in 10 infants, two pathogens were identified. The mean gestational age (GA) of the infants with LOS was 33.2±4.8 weeks (23 to 42 weeks). Gram-positive cocci (GPC) caused most of the LOS episodes (65.8%, 50/76), with coagulase-negative Staphylococcus (CoNS) as the most common cause of LOS (50%, 38/76). Gram-negative rod species (GNRs) accounted for 32.9% (25/76) of the LOS cases, and fungi accounted for 1.3% (1/76). The mortality rates for GNR and GPC were 17.9% and 6.4%, respectively (p>0.05). The mean CRP and conjugated bilirubin levels for the GPC and GNR groups were 37.5 vs. 29.5 mg/dl and 0.7 vs 1.5 mg/dl, respectively (p>0.05). GNRs had a 20–25% ceftriaxone resistance. Two (4%) GPC species were resistant to linezolid, while all were susceptible to vancomycin. All of the GNRs were susceptible to carbapenems.

CONCLUSION: These results underscore the recent emergence of CoNS in NICUs. LOS due to GNRs seems to display higher C-reactive protein and conjugated bilirubin values than those due to GPC. Clinical monitoring of NIs and bacterial resistance profiles are required in all NICUs.

Keywords: Antibiotic susceptibility; etiology of neonatal sepsis; newborn; nosocomial infection; sepsis.

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Although life-saving benefits have been shown for intensive medical care, recent advances may expose critically ill patients to both infectious and noninfectious complications. Neonates represent a unique and highly vulnerable patient population because of their immunologic immaturity and altered cutaneous barriers. Thus, newborns in neonatal intensive care units (NICUs) are at high risk for developing nosocomial infections (NIs), which results in increased risk for morbidity and mortality [1]. Neonatal risk factors that have been shown to contribute to neonatal sepsis are prematurity, low birth weight, asphyxia, congenital anomalies, exposure to invasive procedures, and prolonged stays in the NICU [2, 3].

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Despite site-specific infection rates vary by birth weight, NICU NI rates range from 6% to 25% [4–6].

Commonly isolated bacterial species from NICUs are *Klebsiella pneumonia*, *Escherichia coli*, *Enterobacter* spp, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Streptococcus* spp, *Citrobacter* spp, and coagulase-negative *Staphylococcus* (CoNS) [7–9].

Neonatal sepsis may be defined either clinically and/or microbiologically by positive blood and/or cerebrospinal fluid cultures. Also, neonatal sepsis may be classified according to the time of onset of the disease as early-onset sepsis (EOS) and late-onset sepsis (LOS). This distinction has clinical relevance, as EOS is mainly due to bacteria acquired before and during delivery, while LOS is due to bacteria acquired after delivery (from nosocomial or community sources) [9]. In the literature, however, there is little consensus as to what age limits apply. LOS is variably defined as occurring after 72 h in hospitalized infants to after six days in neonates in the community [10].

The bacterial profiles of blood cultures and the bacterial susceptibility patterns in an area can aid decisions on empirical treatment, which are keys of the management of neonatal sepsis. Thus, in the present study, we aimed to ascertain the bacteriological profiles and antimicrobial susceptibility patterns in NIs. The present study aims to give an overview of the burden of bacterial sepsis in the newborn population and provide an objective basis on which empirical antibiotic therapy should be initiated.

### MATERIALS AND METHODS

We conducted a cross-sectional study from December 01, 2014 until January 31, 2016 in the NICU of the Hospital of Tokat Gaziosmanpasa University Faculty of Medicine, Tokat, Turkey. We defined LOS as a positive blood culture after the first three days of life (≥72 h) [10]. We evaluated the confirmed LOS cases retrospectively. All neonates participated had a thorough clinical evaluation and have processed blood cultures for microbiologic analysis. Each specimen has been inoculated directly into a Bact/ALERT (BioMerieux, Laval, Quebec, Canada) pediatric isolator tube. Antibiotic susceptibility, C-reactive protein (CRP), white blood cell, absolute neutrophil count, immature/total neutrophil ratio, platelet count, mean platelet volume, and biochemical parameters like conjugated and unconjugated bilirubin levels were recorded. Additionally, symptoms, demographics (birth weight, gestational age and sex), duration of hospital stay, and outcome data were collected and analyzed. The only outcome recorded was survival.

Only the first episode of NI was included for analysis, and the detected pathogenic organisms were categorized into three clinically relevant groups as follows: gram-negative rods (GNRs), gram-positive cocci (GPC), and fungi.

### Statistical Analysis

Data were obtained regarding the arithmetic mean, standard deviation, frequency, or percentage. Independent samples t-tests were used to compare the variables between groups. A P-value <0.05 was considered statistically significant. Analyses were performed using SPSS 19 (IBM SPSS Statistics 19, SPSS, Somers, NY, USA).

### RESULTS

During the study period, there were 1210 infants in the NICU, and 76 (6.3%) of these infants were noted as having LOS. A total of 86 episodes of LOS were documented; in 10 infants, two pathogens were identified. The mean birth weight of the infants with LOS was 2044±1031 grams (525 to 4435 g). The mean gestational age (GA) of the infants with LOS was 33.2±4.8 weeks (23 to 42 weeks). The cesarean section ratio for all LOS cases was 67.1%. In the first two weeks of life, 56.6% of the infants had a central venous catheter.

GPCs were determined to be the cause of most episodes of LOS (65.8%, 50/76). GNRs accounted for 32.9% (25/76) of LOS cases, and fungi accounted for

### Table 1. Microorganisms isolated from blood cultures

<table>
<thead>
<tr>
<th>Microorganism</th>
<th>%</th>
</tr>
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<tbody>
<tr>
<td>Gram-positive</td>
<td>65.8</td>
</tr>
<tr>
<td>CONS</td>
<td>50</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>10.5</td>
</tr>
<tr>
<td><em>Enterococcus spp</em></td>
<td>4</td>
</tr>
<tr>
<td><em>Streptococcus pneumonia</em></td>
<td>1.3</td>
</tr>
<tr>
<td>Fungus</td>
<td>1.3</td>
</tr>
<tr>
<td><em>Candida spp</em></td>
<td>1.3</td>
</tr>
<tr>
<td>Gram-negative</td>
<td>32.9</td>
</tr>
<tr>
<td><em>Klebsiella spp</em></td>
<td>11.8</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>9.2</td>
</tr>
<tr>
<td><em>Serratia marcescens</em></td>
<td>6.6</td>
</tr>
<tr>
<td><em>Enterobacter</em> spp</td>
<td>4</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>1.3</td>
</tr>
</tbody>
</table>

CONS: Coagulase-negative staphylococcus.
1.3% (1/76). Table 1 shows the distribution of microorganisms that were isolated from blood cultures. The postnatal age at the time of diagnosis was 16.2±11.5 days in the GPC group and 15.4±10.8 days in the GNR group (p>0.05). Central nervous system infection was determined in two cases and urinary tract infection was determined in six cases. Eight of the 76 LOS cases (10.5%) died, and the mortality rates for the GNR and GPC groups were 17.9% and 6.4%, respectively (p>0.05).

The mean CRP levels were 29.5±33.4 and 43.2±54.6 mg/dl for the GPC and GNR groups, respectively (p>0.05). The mean conjugated bilirubin level was 0.7 mg/dl in the GPC group, whereas it was 1.5 mg/dl in the GNR group, as shown in Table 2 (p>0.05).

Skin alterations, such as chilly periphery or cutis marmorata and reduced movement at a rate of 50%, were the most frequent clinical findings. The distribution of the major clinical findings is shown in Table 3.

GNRs showed 20–25% ceftriaxone resistance. Table 4 shows the selected antibiotic susceptibility of GNR- and GPC-positive cases. All were sensitive to carbapenems. Additionally, all GPCs were sensitive to vancomycin. During the study period, vancomycin-resistant *Enterococcus* colonization was present in five infants in the NICU who were all transferred to our NICU from other institutions. None progressed to vancomycin-resistant infection.

**DISCUSSION**

NIs are a major cause of morbidity in NICUs. NIs are associated with lower birth weight and increased thera-
peutic intensity and result in prolonged hospitalization, leading to increased costs [5]. The increasing number of technology-dependent infants is the primary factor leading to the increase of NIs [11]. Although immunologic immaturity and altered cutaneous barriers play a role in the vulnerability of neonates to NIs, therapeutic interventions that are lifesaving for these fragile infants also appear to be associated with the majority of infectious complications resulting in neonatal morbidity and mortality [1, 2].

The prevalence of NIs in NICUs differs among countries and NICUs of the same country. Additionally, seasonal bacteriologic profile changes are likely to occur. Multicenter studies in Europe have reported an overall NICU NI rate of 7% [12]. Consistent with the present study, in Turkey, the neonatal sepsis frequency ranges from 2.1% to 17% in NICUs [7]. Retrospectively evaluation of data-base of 242 neonatal departments has shown the LOS rate in NIs for VLBW infants was 8.1% with 6.4% case fatality risk [13].

The pathogens most often implicated in neonatal sepsis in developing countries differ from the pathogens seen in developed countries. However, GNRs are overall more commonly found to cause LOS in resource-limited countries’ NICUs [8, 14–17]. Consistent with the present study, various authors have reported that LOS mainly arises from GPCs, especially CoNS [16, 18, 19]. Similar to the present study, the Australasian Study Group for Neonatal Infections reported that CoNS predominated and was responsible for 57.1% of the total LOS cases [19–21]. Also, Piening et al. [13] and Wu et al. [22] have reported that the most frequently identified organism was CoNS with the frequency of 56.2% and 40%, respectively. Also, Piening et al. have not reported any case of isolation of Group B Streptococcus. The reasons underlying the rise of CoNS sepsis are likely to be multifactorial, attributable in part to invasive procedures, the thin, fragile skin of neonates, and the widespread use of central venous catheters and lipid emulsions in parenteral nutrition [20, 23]. According to the low mortality rate, CoNS infections are relatively benign infections, at least in comparison with GNR or S. aureus infections [16, 24]. In consistency with our study, it has shown that a more than three-fold increased adjusted case fatality risk caused by gram-negative bacteria compared to the GPC, especially Staphylococcus species [13]. Despite being the most frequently isolated pathogen of neonatal sepsis, the significance of CoNS in neonatal morbidities may be underestimated and needs to be elucidated [25].

Although our results did not reach statistical significance, CRP and conjugated bilirubin tended to be higher and platelet levels tended to be lower in GNR sepsis. The lack of statistical significance could be due to the small sample size in this study. The results from this study further add to the evidence that early-onset cholestatic jaundice is common in GNR neonatal septicemia. Khalil et al. [26] reported that cholestatic jaundice occurred at a rate of 42.5% in GNR septicemia [26]. In a retrospective analysis of neonatal sepsis cases, 25% of the infants had some degree of cholestasis [6]. Cholestasis may be due to the pathogenic role of circulating endotoxins in causing biliary stasis and hepatic parenchymal injury [26].

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>Gram-negative</th>
<th>Gram-positive</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Tested isolates</td>
<td>Susceptible</td>
</tr>
<tr>
<td></td>
<td>n</td>
<td>n</td>
</tr>
<tr>
<td>Ceftriaxone</td>
<td>9</td>
<td>7</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>21</td>
<td>16</td>
</tr>
<tr>
<td>Linezolid</td>
<td>44</td>
<td>96</td>
</tr>
<tr>
<td>Meropenem</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>Piperacillin-Tazobactam</td>
<td>8</td>
<td>7</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>12</td>
<td>9</td>
</tr>
<tr>
<td>Cefepime</td>
<td>14</td>
<td>13</td>
</tr>
</tbody>
</table>

Table 4. Selected antibiotic susceptibility of gram-negative and gram-positive microorganisms
Although all GNRs were susceptible to carbapenems, the sensitivity to ceftriaxone was 78% in the present study. Cephalosporin resistance in GNRs is an increasing problem in NICUs that makes challenging the decision of empiric antibiotic treatment [27].

LOS due to GNRs seems to display higher CRP and conjugated bilirubin values than cases that arise from GPCs. To accelerate the pace toward improved outcomes, we must first develop and validate an accepted consensus on bacteriological profiles and their antimicrobial susceptibility patterns for every NICU. Simple clues in clinical and laboratory findings may help to estimate whether the bacteria gram-positive or negative before the blood culture results. For best results in infection management, nurseries should periodically review their bacterial sensitivity patterns and antibiotic policies.

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